# organic papers

Acta Crystallographica Section E Structure Reports Online

ISSN 1600-5368

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#### Key indicators

Single-crystal X-ray study  $T = 183 K$ Mean  $\sigma$ (C–C) = 0.005 Å  $R$  factor = 0.046  $wR$  factor = 0.154 Data-to-parameter ratio = 10.7

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# Benzyl 2-deoxy-2-(3,5-di-tert-butylsalicylamino) *a*-D-glucopyranoside

The title compound,  $C_{28}H_{41}NO_6$ , was obtained by reduction of benzyl 2-deoxy-2-(3,5-di-tert-butylsalicylideneamino)-α-Dglucopyranoside. The benzyl substituent is in an axial position, whereas the 3,5-di-tert-butylsalicylamino substituent and the hydroxyl groups are in equatorial positions. The enantiomerically pure title compound is a potential  $O, N, O$ -chelate ligand suitable as a precursor for chiral transition metal complexes. The absolute configuration was determined by NMR experiments.

## Comment

Related imino-functionalized carbohydrates containing an unprotected anomeric hydroxyl group are known as chiral chelate ligands. The cis-VO<sub>2</sub><sup>+</sup>, cis-MoO<sub>2</sub><sup>2+</sup> and trans-UO<sub>2</sub><sup>2+</sup> complexes are effective catalysts in asymmetric olefin epoxidation (Zhao et al., 2003; Sah et al., 2001) or sulfide oxidation (Cucciolito et al., 2005). Tc and Re complexes have been developed for molecular imaging and radiotherapy (Duatti et al., 1987; Bayly et al., 2004). The reduced amino-derived compounds are stable towards hydrolysis. The title compound, (I), has been synthesized as part of our efforts to utilize sugarmodified Schiff base ligands (Burkhardt et al., 2007a,b) as precursors for chiral catalysts (Becher et al., 2006) and chiral polynuclear complexes of paramagnetic metal centers (Burkhardt et al., 2006; Roth et al., 2006).



The hydroxyl group of the anomeric atom C1 is blocked via a benzyl ether. The benzyl substituent is in an axial position, corresponding to the exclusive presence of the  $\alpha$ -anomer as confirmed by <sup>1</sup>H NMR data. H1 leads to a doublet with  ${}^{3}J_{1,2}$  = 3.7 Hz associated with the cis configuration of the atoms H1

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Received 30 January 2007 Accepted 14 May 2007



### Figure 1

A view of the molecular structure of the title compound. Displacement ellipsoids are drawn at the 50% probability level.



#### Figure 2

Supramolecular double-chain of  $C_{28}H_{41}NO_6$ . Hydrogen bonds are shown as dashed lines and H atoms have been omitted.

and H2. All other substituents at C2–C5 are in equatorial positions.

The 3,5-di-tert-butylsalicylamino group at C2 was introduced via a three-step-synthesis into the sugar backbone. Alkaline hydrolysis of 2-acetamido-2-deoxy- $\alpha$ -D-glucopyranoside yielded the amino sugar and, by reaction with 3,5-di-tertbutyl-salicylaldehyde, the Schiff base was formed. The imine was finally reduced with NaBH<sub>4</sub> to the title compound, which is soluble in  $CHCl<sub>3</sub>$  and several alcohols.

The sugar ring in (I) adopts the more stable  ${}^{4}C_1$  chair conformation (see Fig. 1). The benzyl group at C1 is directed towards the hydroxyl group at C6. The mean plane of the anomeric substituent (defined by C8–C13) and the mean sugar plane (defined by C1, C3, C4 and O1) subtend an angle of 57.03  $(12)^\circ$ . The mean plane of the 3,5-di-tert-butylsalicylamino substituent at C2 (defined by C15–C20) and the mean sugar plane subtend an angle of  $81.68$   $(11)^\circ$  accompanied with the nearly perpendicular orientation of the substituent's aromatic ring towards the carbohydrate chair.

The individual  $C_{28}H_{41}NO_6$  units are linked *via* intermolecular hydrogen bonding, forming one-dimensional double chains with a twofold screw symmetry along the crystallographic  $a$  axis (see Fig. 2). The hydroxyl groups at C4 and C6 of the monosaccharide groups are involved in the hydrogen-bonding network.

The title compound is suitable as an O,N,O-chelate ligand. Complexation in protic solvents should work better than with the corresponding Schiff base ligands because of the hydrolysis stability of the secondary amine in comparison with imine in the Schiff base.

## Experimental

All substances were purchased from commercial suppliers and used without further purification. Benzyl 2-acetamido-2-deoxy- $\alpha$ -Dglucopyranoside was prepared from  $N$ -acetyl- $\alpha$ -D-glucoseamine according to Györgydéak (1991).

Compound (I) was obtained by reduction of benzyl 2-deoxy-2- (3,5-di-tert-butylsalicylideneamino)-a-D-glucopyranoside.

For the preparation of benzyl 2-deoxy-2-(3,5-di-tert-butylsalicylideneamino)- $\alpha$ -D-glucopyranoside, a solution of benzyl 2-acetamido- $2$ -deoxy- $\alpha$ -D-glucopyranoside  $(2.00 \text{ g}, 0.006 \text{ mol})$  and KOH  $(14.00 \text{ g},$ 0.250 mol) in 50 ml of ethanol (96%) was refluxed overnight. After cooling to 298 K, the orange mixture was diluted with 100 ml of ethanol (96%) and neutralized with HCl. The precipitated KCl was separated by centrifugation and the filtrate volume was reduced in vacuo to 10 ml. Addition of NaHCO<sub>3</sub> (700 mg, 8.33 mmol) and 3,5-ditert-butyl-salicylaldehyde (1.52 g, 0.006 mol) afforded an intensely yellow mixture. After 2.5 h of stirring at 298 K, a pale-yellow precipitate was obtained. The amorphous solid was collected by filtration and washed successively with 100 ml of water and 100 ml of n-hexane with vigorous stirring for 12 h each. After filtration the product was dried in vacuo at 333 K. Yield 2.09 g (67%). M.p. 498–500 K (water). IR (KBr): 3494, 3401, 3276 ( $\nu$  O—H), 3066, 3027 ( $\nu$  C—H arom.), 2962 ( $v_{\text{as}}$  CH<sub>3</sub>), 2870 ( $v_{\text{s}}$  CH<sub>2</sub>), 1628 ( $v$  CH=N), 1597, 1498 ( $v$  C=C), 1469, 1455, 1441 ( $\delta_{as}$  CH<sub>3</sub>,  $\delta$  CH<sub>2</sub>), 1392, 1362 ( $\delta_{s}$  CH<sub>3</sub>), 1335, 1307,  $1271, 1251, 1207, 1158, 1135, 1083, 1043, 1024, 1007, 983 (v C - O),$ 880, 850, 827, 806, 772, 739, 731, 712, 696, 645, 530, 514 cm<sup>-1</sup>; <sup>1</sup>H NMR  $(400 \text{ MHz}, \text{DMSO-}d_6, 298 \text{ K}): 1.27 \text{ and } 1.43 \text{ (2s, each 9H, H methyl)},$ 3.22–3.28 (m, 1H, H4), 3.26 (dd,  ${}^{3}J_{21} = 3.6$  Hz,  ${}^{3}J_{23} = 9.5$  Hz, 1H, H2), 3.54–3.64 (m, 2H, H5 and H6A), 3.69–3.71 (m, 2H, H3 and H6B), 4.51 and 4.74 (2d,  $^{2}J_{7A7B}$  = 12.4 Hz, each 1H, H7A and H7B), 4.60 (dd,  $^{3}J_{\text{av}}$  $=$  5.9 Hz, 1H, OH6), 4.86 (d,  $^{3}J_{12} = 3.7$  Hz, 1H, H1), 5.05 (d,  $^{3}J_{4OH4} =$ 5.9 Hz, 1H, OH4), 5.11 (d,  $3J_{3OH3} = 5.9$  Hz, 1H, OH3), 7.25–7.32 (m, 3H, H ph), 7.41–7.43 (m, 2H, H ph), 8.49 (s, 1H, H14), 14.70 (s, 1H, OH20) p.p.m; <sup>13</sup>C NMR (60 MHz, DMSO- $d_6$ , 298 K): 29.2 and 31.3 (C22–C24 and C26–C28), 33.8 and 34.6 (C21 and C25), 60.9 (C6), 67.9 (C7), 70.2 and 71.4 (C2, C3 and C4), 73.4 (C5), 97.5 (C1), 117.6, 126.1, 126.2, 127.4, 128.0, 135.8, 137.9 and 139.0 (C ph), 158.6 (C20) 168.1 (C14) p.p.m. ESI–MS:  $m/z$  (%) = 486 [M+H]<sup>+</sup> (3), 508 [M+Na]<sup>+</sup> (100).  $C_{28}H_{39}NO_6$  (485.61 g mol<sup>-1</sup>): calculated C 69.25, H 8.09, N 2.88%; found C 69.10, H 8.17, N 2.68%.

For the preparation of benzyl 2-deoxy-2-(3,5-di-tert-butylsalicylamino)- $\alpha$ -D-glucopyranoside, with ice-cooling, a solution of benzyl 2-deoxy-2-(3,5-di-tert-butylsalicylideneamino)-a-D-glucopyranoside (0.50 g, 0.001 mol) in 25 ml of ethanol (100%) was slowly poured into a suspension of NaBH<sub>4</sub> (0.20 g, 0.005 mol) in 10 ml of ethanol (100%). The slightly yellow solution was stirred for 5 h at 298 K. The reaction was quenched by addition of  $1 M$  HCl until pH 5 was achieved. After complete removal of the solvent, the pale-brown semi-solid residue was taken up in 70 ml of CHCl<sub>3</sub>. The organic layer was washed once with an  $NaHCO<sub>3</sub>$  solution (pH 8) and four times with water, and finally dried over  $Na<sub>2</sub>SO<sub>4</sub>$ . Filtration and complete removal of the solvent yielded the raw product as a colourless amorphous solid which was dried in vacuo at 333 K. Single crystals suitable for X-ray determination were obtained by slow evaporation of a solution of the crude product in 15 ml of methanol over a period of six weeks. Yield 320 mg (64%). M.p. 465–467 K (methanol). IR (KBr): 3513 ( $\nu$  O-H), 3324 ( $\nu$  N-H), 3031, 3000 ( $\nu$  C-H arom.), 2954 ( $v_{\rm as}$  CH<sub>3</sub>), 2909 ( $v_{\rm as}$  CH<sub>2</sub>), 2870 ( $v_{\rm s}$  CH<sub>2</sub>), 2835 ( $v_{\rm s}$  CH<sub>3</sub>), 1481, 1456 (δ<sub>as</sub> CH<sub>3</sub>, δ CH<sub>2</sub>), 1360 (δ<sub>s</sub> CH<sub>3</sub>), 1238, 1127, 1096, 1063, 987 (ν C– O), 763, 702 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ , 298 K): 1.22 and 1.35 (2s, 18H, H methyl), 2.49–2.53 (m, 1H, H2), 2.73 (d,  $3J = 5.4$  Hz, 1H, N–H), 3.13  $(dd, {}^{3}J_{43} = {}^{3}J_{45} = 9.3$  Hz, 1H, H4), 3.41–3.50  $(m, 3H,$ H3, H5 and H6), 3.66  $(dd, {}^3J_{65} = {}^2J_{6a6e} = 11.3$  Hz, 1H, H6), 3.81–3.86  $(m, 2H, H14)$ , 4.46 and 4.67  $(2d, {}^{2}J_{7a7b} = 11.7 \text{ Hz}$ , each 1H, H7A and H7B), 4.53 (s, 1H, OH6), 4.95 (s, 1H, OH4), 4.98 (d,  $^{3}J_{12} = 3.7$  Hz, 1H, H1), 5.02 (s, 1H, OH3), 6.80 (d,  $J = 2.2$  Hz, 1H, H ph), 7.07 (d,  $J =$ 2.2 Hz, 1H, H ph), 7.28–7.36 (m, 3H, H ph), 7.45–7.47 (m, 2H, H ph) p.p.m.; <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ , 298 K): 29.5 and 31.5 (C22– C24 and C26–C28), 33.7 and 34.4 (C21 and C25), 50.8 (C14), 60.9 (C6), 61.1 (C2), 68.2 (C7), 70.5 (C4), 72.2 and 73.1 (C3 and C5), 95.4 (C1), 121.6, 122.9, 123.0, 127.5, 128.1, 128.2, 134.6, 137.7 and 139.3 (C ph), 154.4 (C20) p.p.m. ESI-MS:  $m/z$  (%) =  $[M+H]^+$  (4),  $[M+Na]^+$ (40).  $C_{28}H_{41}NO_6$  (487.63 g mol<sup>-1</sup>): calculated: C 68.97, H 8.47, N 2.87%; found: C 69.09, H 8.40, N 2.72%.

 $V = 2700.0$  (9)  $\AA^3$ 

Mo  $K\alpha$  radiation  $\mu = 0.08$  mm<sup>-1</sup>  $T = 183$  (2) K  $0.7 \times 0.7 \times 0.4$  mm

 $R_{\text{int}} = 0.072$ 

3554 independent reflections 2415 reflections with  $I > 2\sigma(I)$ 

 $Z = 4$ 

#### Crystal data



Data collection

Nonius KappaCCD diffractometer Absorption correction: none 18295 measured reflections

### Refinement



C-bound H atoms were positioned geometrically  $[C-H = 0.95]$  $(Csp<sup>2</sup>)$ , 0.98 (methyl), 0.99 (methylene) and 1.00 Å (methine), and

 $O-H = 0.84$  Å and treated as riding atoms with fixed displacement parameters  $[U_{iso}(H) = xU_{eq}(C)$ , where  $x = 1.5$  for methyl and OH groups and 1.2 for all others]. The H atom on the N atom was located and refined with an isotropic displacement parameter. In the absence of significant anomalous scattering effects, Friedel equivalents were merged prior to the final refinement. The absolute configuration was assigned by reference to the chiral starting material and the evidence provided by NMR spectroscopy.

Data collection: COLLECT (Nonius 1998); cell refinement: DENZO (Otwinowski & Minor, 1997); data reduction: DENZO; program(s) used to solve structure: SHELXS97 (Sheldrick, 1997); program(s) used to refine structure: SHELXL97 (Sheldrick, 1997); molecular graphics: XP (Siemens, 1990); software used to prepare material for publication: XP and SHELXL97.

We gratefully thank the Deutsche Forschungsgemeinschaft for financial support.

#### References

- Bayly, S. R., Fisher, C. L., Storr, T., Adam, M. J. & Orvig, C. (2004). Bioconjugate Chem. 15, 923–926.
- Becher, J., Seidel, I., Plass, W. & Klemm, D. (2006). Tetrahedron, 62, 5675– 5681.
- Burkhardt, A., Buchholz, A., Görls, H. & Plass, W. (2006). Eur. J. Inorg. Chem. pp. 3400–3406.
- Burkhardt, A., Buchholz, A., Görls, H. & Plass, W. (2007a). Acta Cryst. E63, o384–o386.
- Burkhardt, A., Buchholz, A., Görls, H. & Plass, W. (2007b). Acta Cryst. E63, o387–o388.
- Cucciolito, M. E., Litto, R. D., Roviello, G. & Ruffo, F. (2005). J. Mol. Catal. A Chem. 236, 176–181.
- Duatti, A., Marchi, A., Magon, L., Deutsch, E., Bertolasi, V. & Gillis, G. (1987). Inorg. Chem. 26, 2182–2186.
- Györgydéak, Z. (1991). Liebigs Ann. Chem. pp. 1291-1300.
- Nonius (1998). COLLECT. Nonius BV, Delft, The Netherlands.
- Otwinowski, Z. & Minor, W. (1997). Methods in Enzymology, Vol. 276, Macromolecular Crystallography, Part A, edited by C. W. Carter Jr & R. M. Sweet, pp. 307–326. New York: Academic Press.
- Roth, A., Becher, J., Herrmann, C., Görls, H., Vaughan, G., Reiher, M., Klemm, D. & Plass, W. (2006). Inorg. Chem. 45, 10066–10076.
- Sah, A. K., Rao, C. P., Saarenketo, P. K., Wegelius, E. K., Kolehmainen, E. & Rissanen, K. (2001). Eur. J. Inorg. Chem. pp. 2773–2781.
- Sheldrick, G. M. (1997). SHELXS97 and SHELXL97. University of Göttingen, Germany.
- Siemens (1990). XP. Version 4.2. Siemens Analytical X-ray Instruments Inc., Karlsruhe, Germany.
- Zhao, J., Zhou, X., Santos, A. M., Herdtweck, E., Romao, C. C. & Kühn, F. E. (2003). Dalton Trans. pp. 3736–3742.